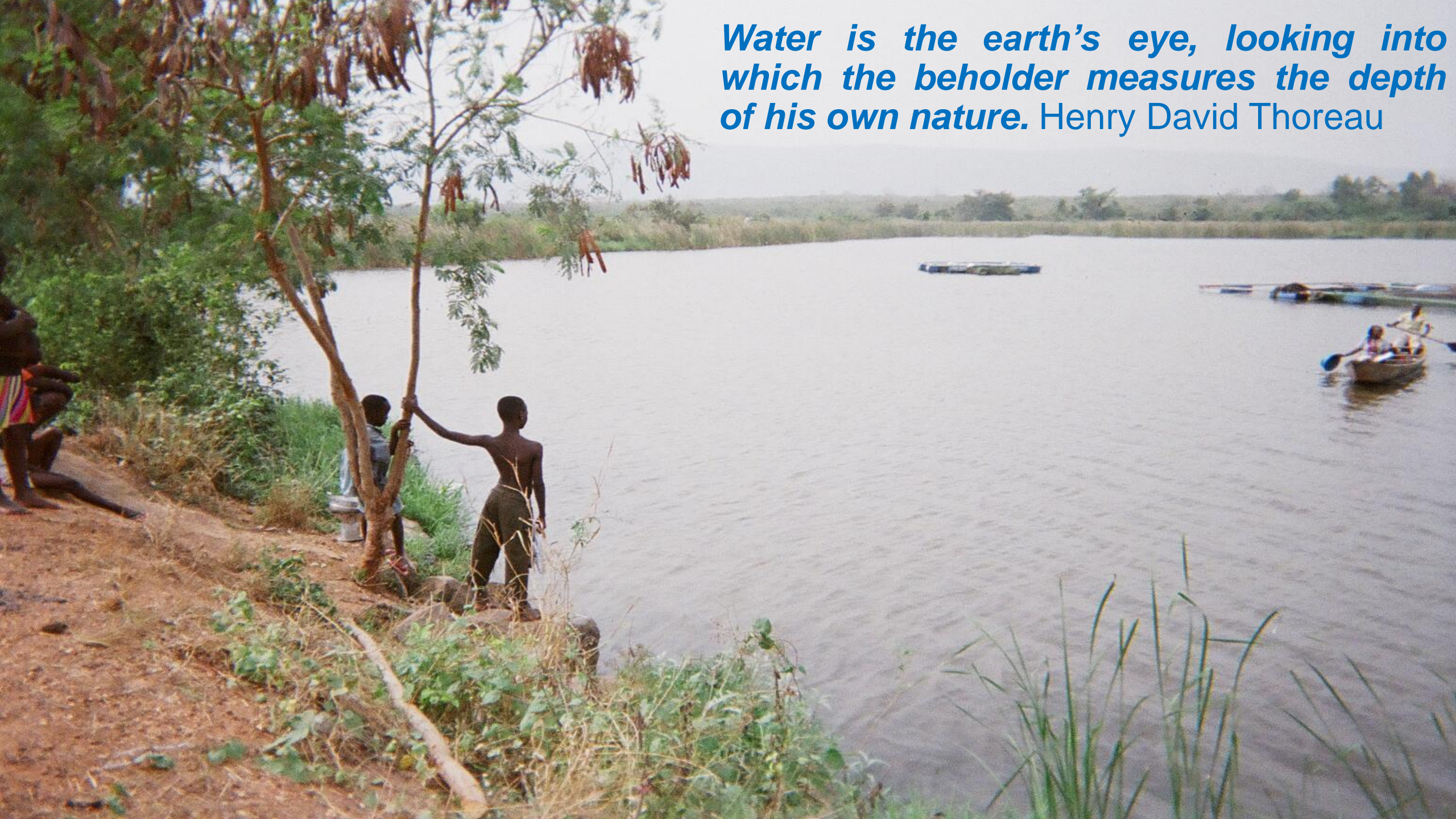


Water is the earth's eye, looking into which the beholder measures the depth of his own nature. Henry David Thoreau





Biological monitoring of coastal waters

Lailah Gifty Akita

lailah.akita@gmail.com

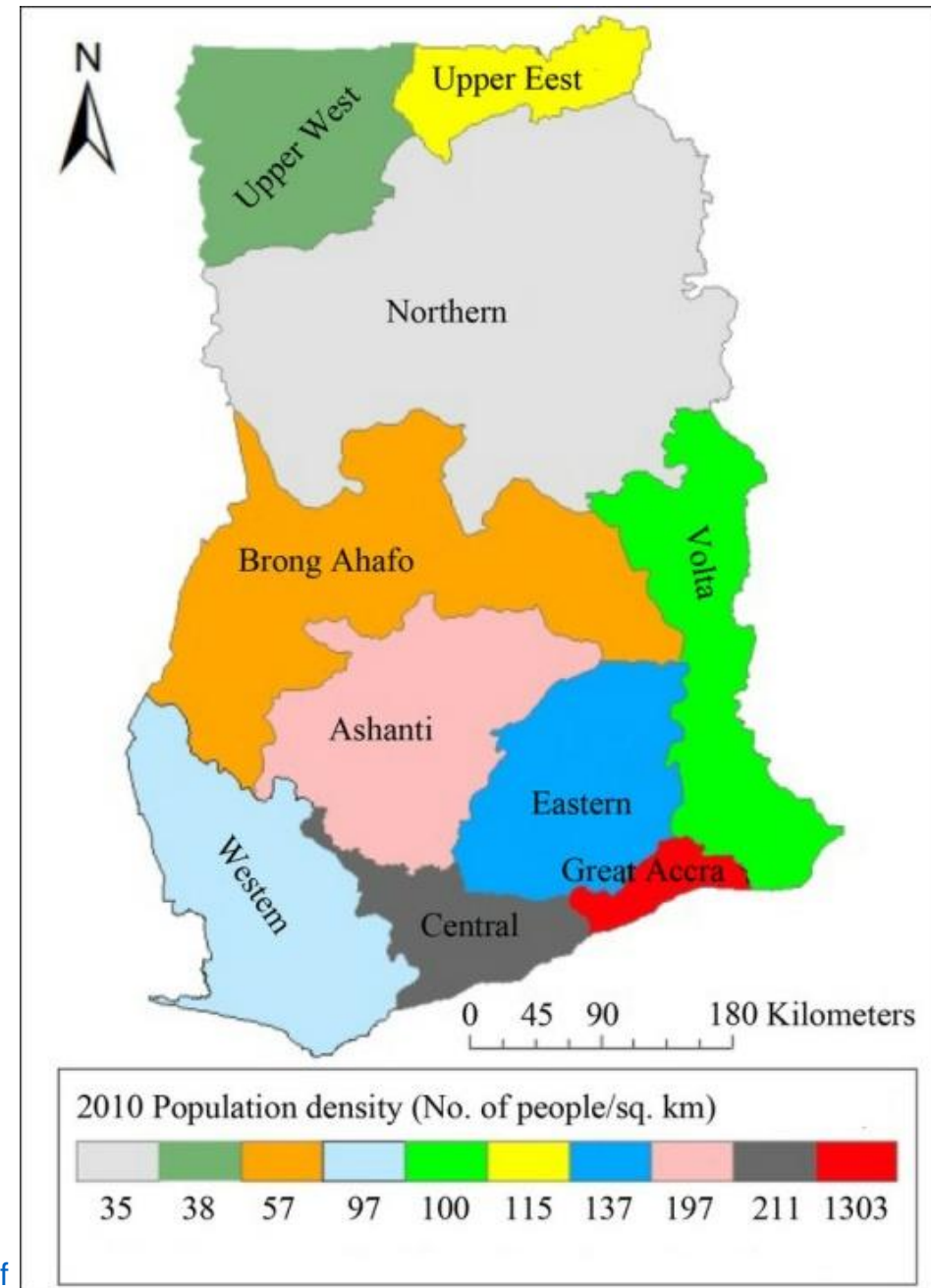
African Initiative of the Volkswagen Foundation

Ghana Summer School on Coastal Environment ,1-5 August 2016

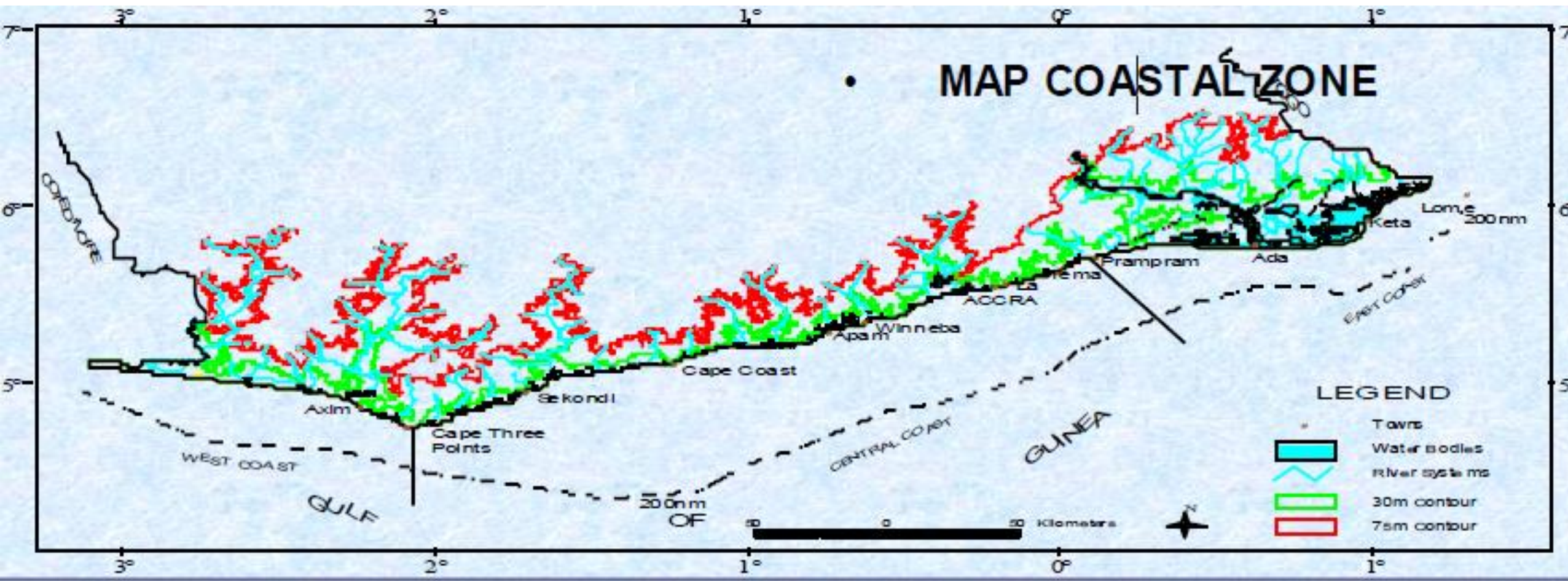
Content

- Background and motivation
- Sampling methods
- Analysis
- Conclusions

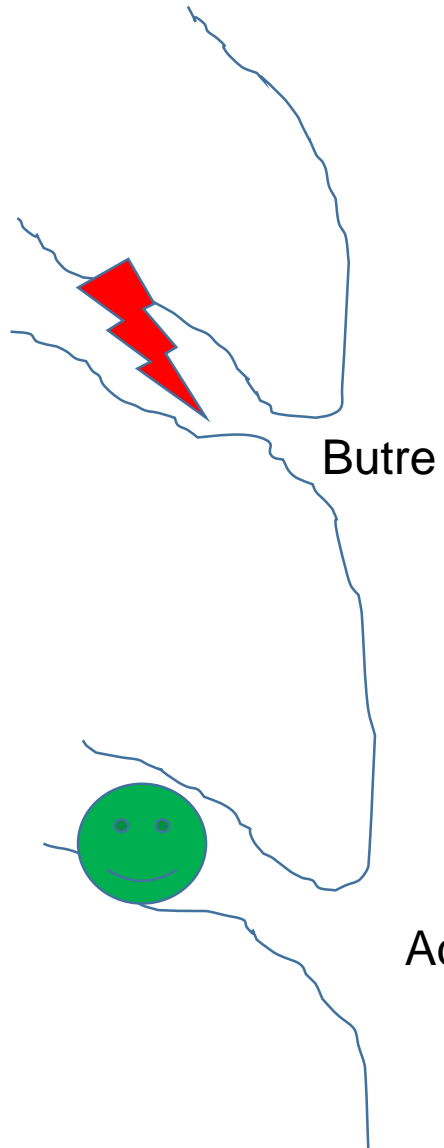
Region: Ghana



Coastal zone of Ghana and sensitivity Mapping

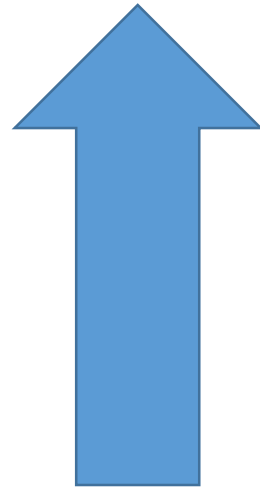


Sampling sites



Butre

Ada



Water quality: Pollution

- **Healthy ecosystems** : maintenance of biodiversity and human well-being.
- **Contamination of water** bodies by chemical, physical, radioactive and pathogenic microbial substances.
- **Global and local concern**: ~ **50 million deaths** per year worldwide (e.g. majority in Africa and Asia).

Source: <http://www.eoearth.org/view/article/156920>

<http://ww2.unhabitat.org/programmes/water/documents/waterreport2.pdf>

Major sources of pollution in Ghana



Solid waste



Pesticides misuse



Sewage outfall



Poisoning the poor – Electronic Waste in Ghana

Effects of pollution

- **Loss of ecosystems** : Species mortality
Biodiversity reduction
Ecosystem services
- **Health risks** : Viruses, bacteria and protozoan
Waterborne diseases
illness and mortality

Need for environmental monitoring

- Supports the **protection of the ecosystems** and **human health** from toxic contaminants.
- Indicate any **alarming environmental** situation.
- Provide opportunities for adoption of any **control measures**.

Need for environmental monitoring:

- Environment science **policy development.**

REVIEWS REVIEWS REVIEWS

Who needs environmental monitoring?

Gary M Lovett^{1*}, Douglas A Burns², Charles T Driscoll³, Jennifer C Jenkins⁴, Myron J Mitchell⁵, Lindsey Rustad⁶, James B Shanley⁷, Gene E Likens¹, and Richard Haeuber⁸

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Environmental monitoring is often criticized as being unscientific, too expensive, and wasteful. While some monitoring studies do suffer from these problems, there are also many **highly successful long-term monitoring programs** that have provided important **scientific advances and crucial information for environmental policy.** Here, we discuss the characteristics of effective monitoring programs, and contend that monitoring should be considered a **fundamental component of environmental science and policy.** We urge scientists who develop monitoring programs to plan in advance to ensure high data quality, accessibility, and cost-effectiveness, and we urge government agencies and other funding institutions to make greater commitments to increasing the amount and **long-term stability of funding for environmental monitoring programs.**

Front Ecol Environ 2007; 5(5): 253–260

In a nutshell:

- Environmental monitoring is often criticized as being unscientific, expensive, and wasteful
- We argue that monitoring is a crucial part of environmental science, costs very little relative to the value of the resources it protects and the policy it informs, and has added value in that basic environmental monitoring data can be used for multiple purposes
- **Effective monitoring** programs address clear questions, use consistent and accepted methods to produce high-quality data, include provisions for management and accessibility of samples and data, and **integrate monitoring into research programs that foster continual examination and use of the data**
- Government agencies should commit to long-term support for valuable monitoring programs, and funders of basic ecological and **environmental research should recognize that monitoring is a fundamental part of environmental science**

Why are benthic organisms important?

1. Important link in the food web - primary producers to higher levels:

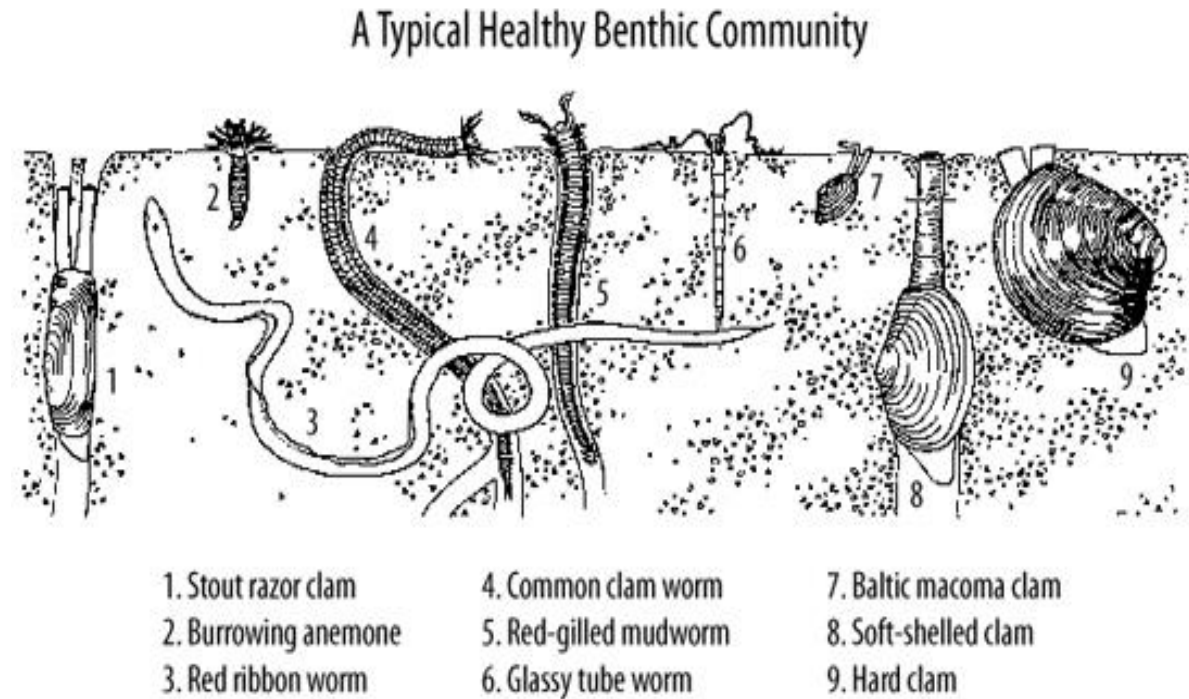
(filter feeder, benthic, detritus feeders)

2. Excellent environmental snapshot - exposure stressors :

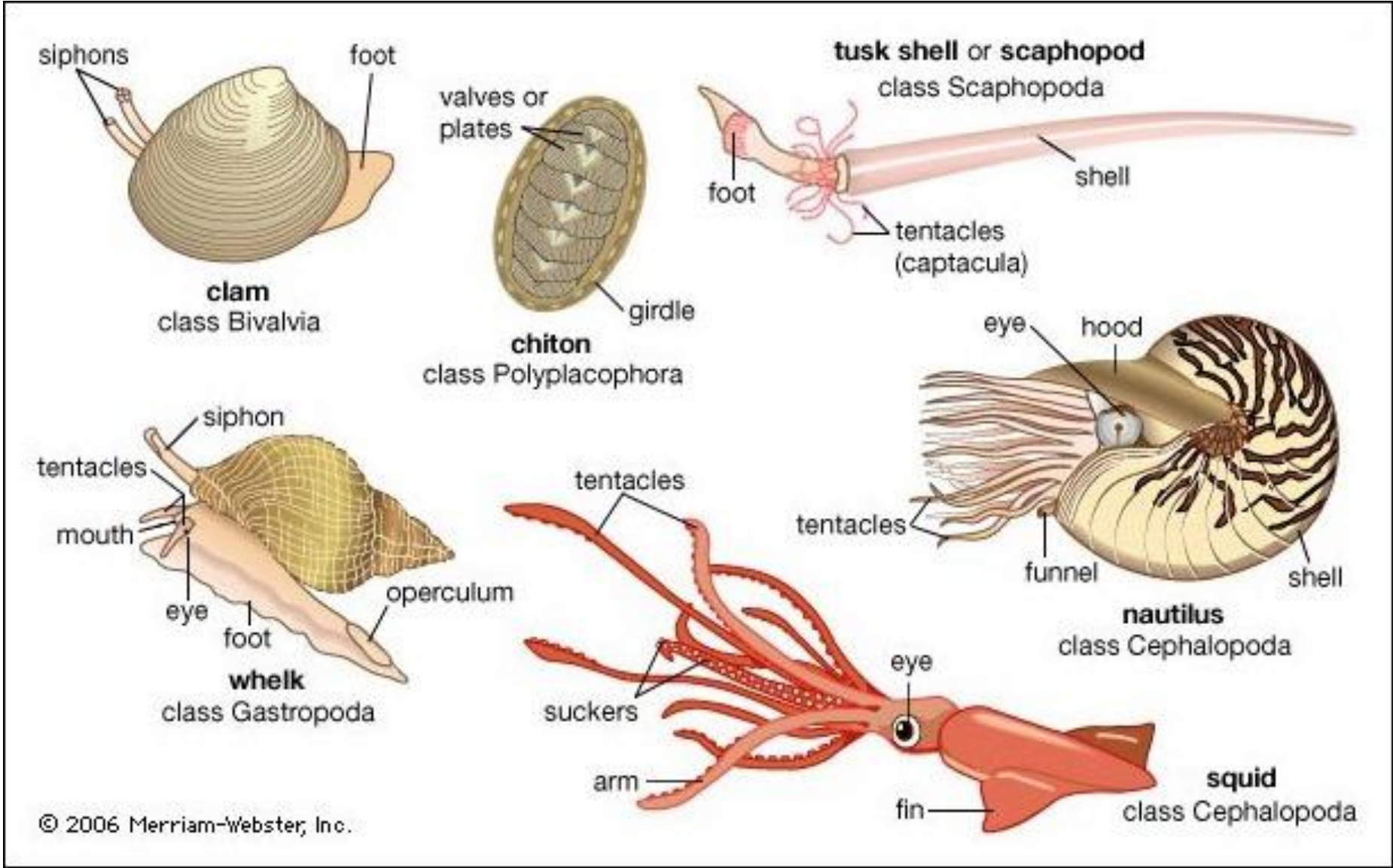
(low oxygen, excess sediment& contaminants)

Benthic Communities

- **Macrofauna**
- **Meiofauna**
- **Preservable hard part: fossil record**
 - ➔ reconstructing natural conditions and identification of pollution events



Identification in mollusks



Source: <http://diet.yukozimo.com/media/o/7219-2.jpg>

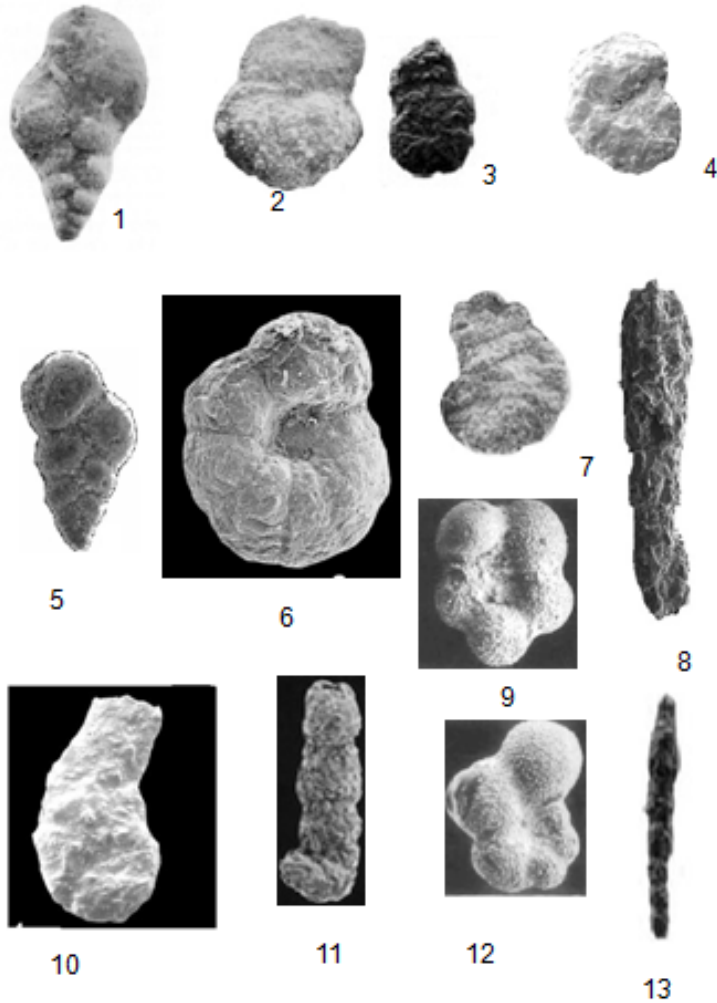
Identification of thecamoebians and ostracods



a) Thecamoebians: 1, *Diffugia corona* Wallich, 1864, 2-12, *Diffugia oblonga* Ehrenberg, 1832, 13-16, *Diffugia tricuspis* Carter, 1865, 17-20, *Diffugia urceolata* Carter, 1864.

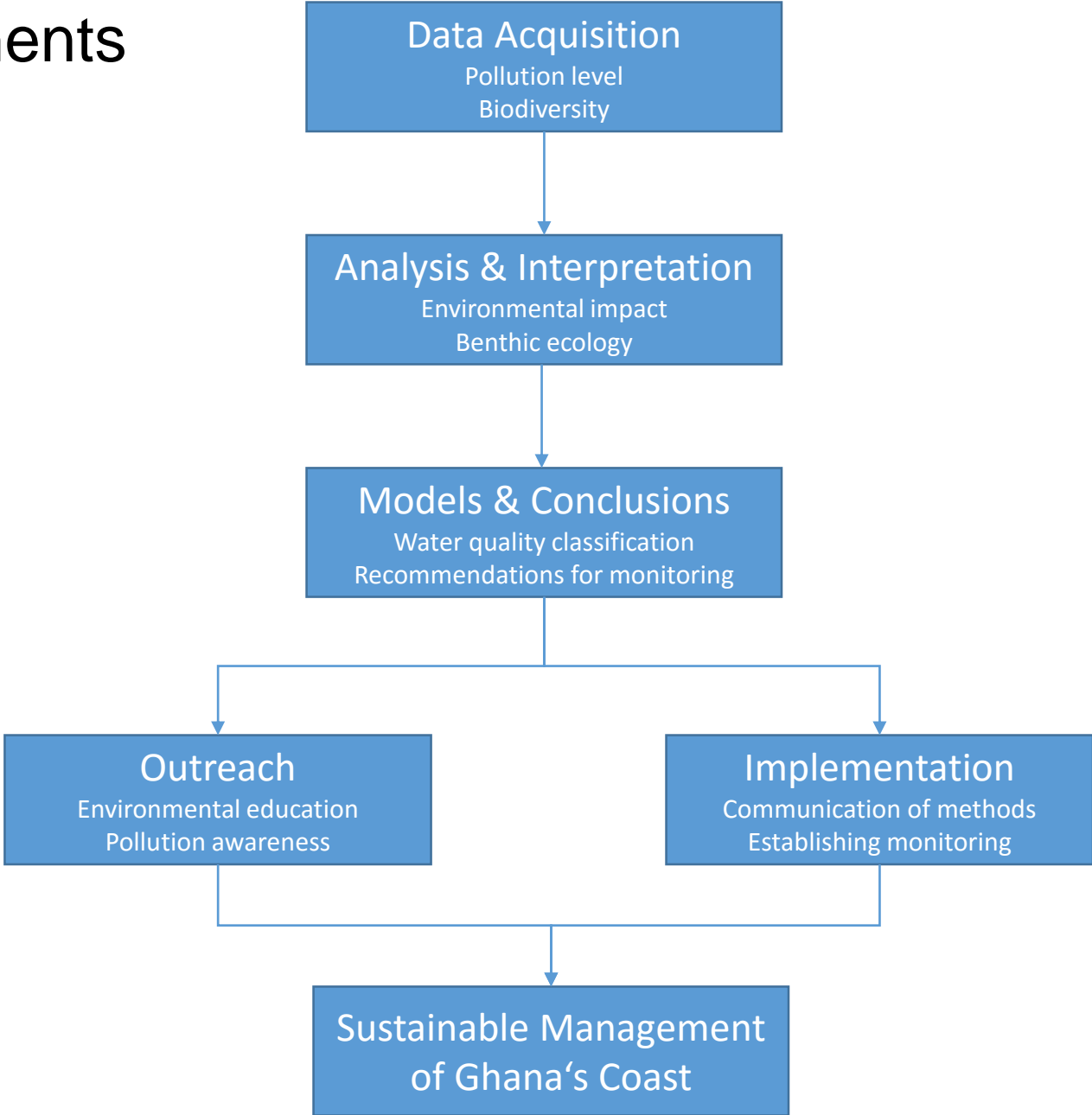
(b) Ostracoda: 21-25, *Afrocythere rostrata* Klie, 1935, 26-28, *Alicennula serricaudata* (Klie, 1935), 30, *Cypria?* sp., 31-33, *Cypridopsis?* sp.

Identification of Foraminifera



1. *Heterohelix reussi*
2. *Ammobaculites numahensis*
3. *Ammobaculites bauchensis*
4. *Haplophragmoides sabariense*
6. *Haplophragmoides* spp.
7. *Ammobaculites pindigensis*
8. *Ammobaculites bauchensis*
9. *Hedbergella* spp.
10. *Ammobaculites pindigensis*
12. *Globigerinelloides* spp.
13. *Reophax guineana*

Project Components



Motivation

Determine

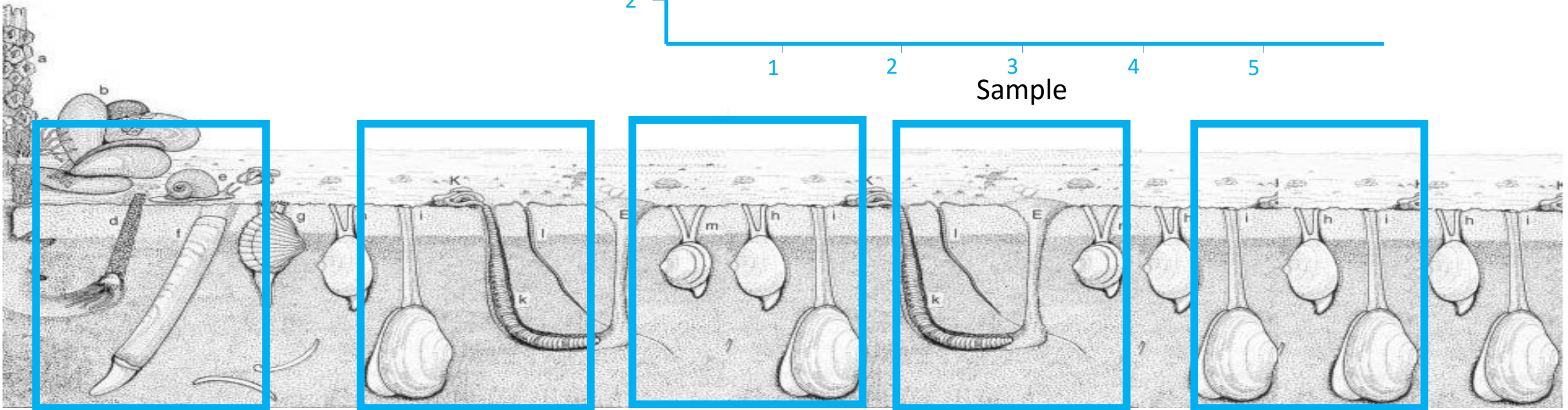
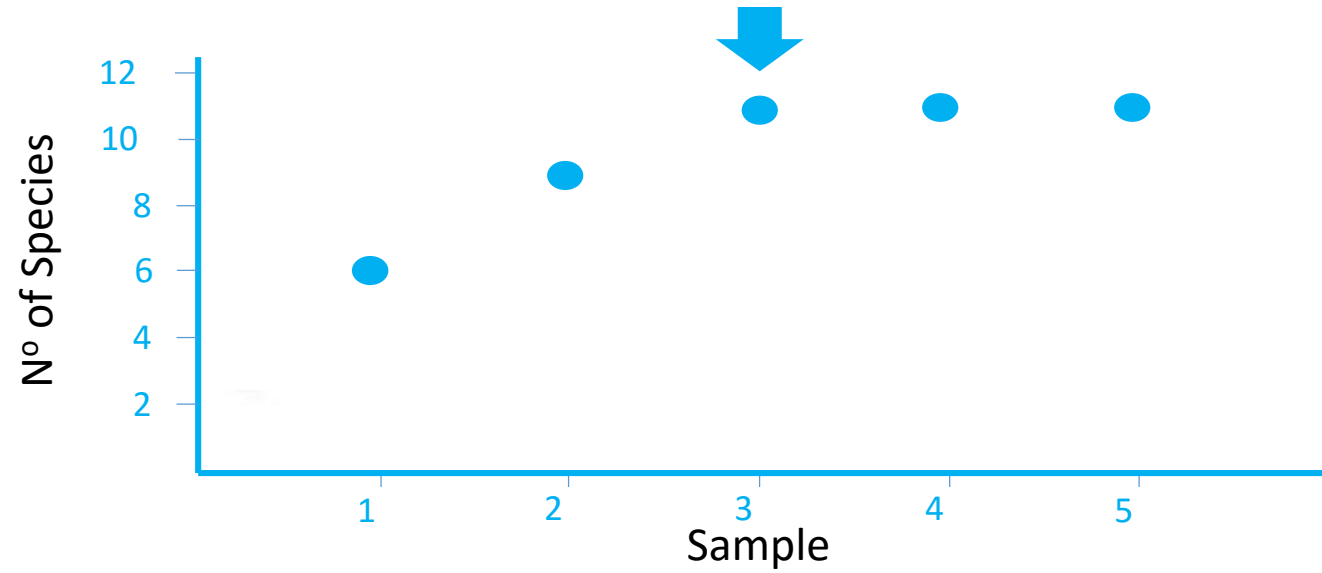
- Species inventories/biodiversity
- Biomass
- Species distribution
- Ecologically driving factors
- Historical data

Sampling

- Assumption: small sample represents large area and complete time interval
- Representative number of replicates

Sampling

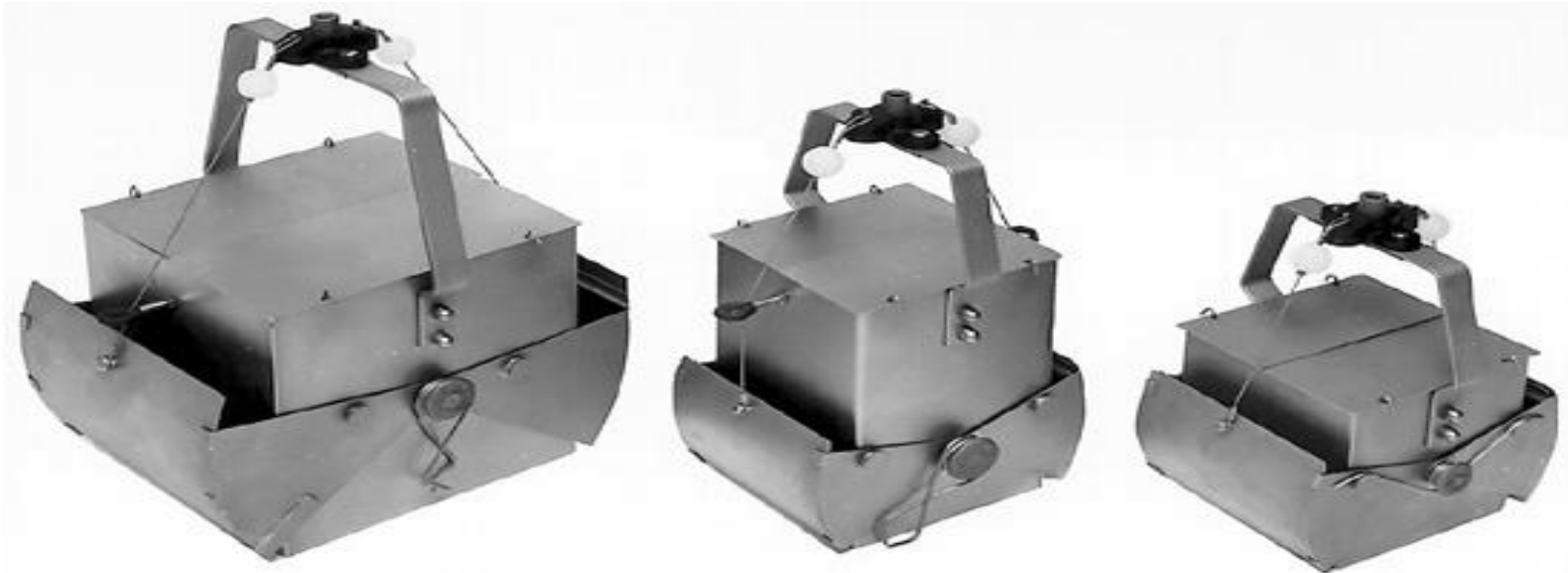
Species-Area curves



Sampling

Box corer sampling

- Penetrates differently according to coarseness of sediment, stones may prevent closing.
- Epibenthic meiofauna and sediment surface not disturbed.



Sampling



- Sieve on **# 0.5 mm** for macrozoobenthos and on **# 0.63 mm + 0.2 mm** for meiobenthos.
- Transfer of **all benthic** in jars.
- Preservation of labelled samples individually in 96% Ethanol, or 4% borax buffered formalin.

Analyses



• Collection

Preservation

Identification
Quantification

Identification/analysis



**Hoosier Riverwatch
Biological Monitoring Data Sheet**

Date ___/___/___ Volunteer ID _____ Site ID _____
 Stream Name _____ Latitude _____ Longitude _____
 Time _____ AM / PM Time Sampling _____ hrs Air Temp _____ C

Current Weather: Clear/Sunny Overcast Showers Rain (steady) Storm (heavy)
 Worst Weather (past 48 hours): Clear/Sunny Overcast Showers Rain (steady) Storm (heavy)

Check Methods Used: Kick Seine Net (3 times) Dip Net (20 jabs or scoops)
 Check Habitats Sampled: Undercut Banks Riffles Leaf Packs Snags/Vegetation Sediment

Pollution Tolerance Index (PTI)

Record the taxa (group) represented in your sampling by either entering the number of organisms you counted or by a

Group 1 - Intolerant	Group 2 - Moderately Intolerant	Group 3 - Fairly Tolerant	Group 4 - Very Tolerant
<input type="checkbox"/> Stonefly Nymph <input type="checkbox"/> Mayfly Nymph <input type="checkbox"/> Caddis Fly Larva <input type="checkbox"/> Riffle Beetle <input type="checkbox"/> Dobsonfly Larva <input type="checkbox"/> Right-Handed Snail <input type="checkbox"/> Water Penny <input type="checkbox"/> # of TAXA <input type="checkbox"/> Weighting Factor (x4)	<input type="checkbox"/> Damselfly Nymph <input type="checkbox"/> Dragonfly Nymph <input type="checkbox"/> Scud <input type="checkbox"/> Sowbug <input type="checkbox"/> Crane Fly Larva <input type="checkbox"/> Clam/Mussels <input type="checkbox"/> Crayfish <input type="checkbox"/> # of TAXA <input type="checkbox"/> Weighting Factor (x3)	<input type="checkbox"/> Leech <input type="checkbox"/> Midge Larva <input type="checkbox"/> Planaria/Flatworm <input type="checkbox"/> Black Fly Larvae <input type="checkbox"/> # of TAXA <input type="checkbox"/> Weighting Factor (x2)	<input type="checkbox"/> Aquatic Worms <input type="checkbox"/> Blood Midge Larva (red) <input type="checkbox"/> Rat-tailed Maggot <input type="checkbox"/> Left-Handed or Pouch Snail <input type="checkbox"/> # of TAXA <input type="checkbox"/> Weighting Factor (x1)

Pollution Tolerance Index Rating

(Add the final index values for each group)

PTI Ratings

Excellent	23 or More
Good	17 - 22
Fair	11 - 16
Bad	10 or Less

Please check other Biological Indicators you observed:
 Native Mussels Zebra Mussels Rusty Crayfish Aquatic Plants _____ %Algae Cover _____ Diversity Index

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Source: <http://midmichigannatureandscience.blogspot.ug/2013/04/aquatic-ecology-and-mother-earth-week.html>

Identification & quantification

		Sample 1	Sample 2	Sample 3	...	PERCENTAGE
FAMILIES	SPECIES	n	n	n	n	%
Actinostolidae	<i>Antholoba achates</i>			1	1	< 0.01
Actinidae	<i>Anemonia alicemartinae</i>	5			5	
Sagartiidae	<i>Anthothoe chilensis</i>	2	2	23	27	0.12
Lineidae	<i>Lineus</i> sp.	7	6	7	20	0.09
Flabelligeridae	<i>Pherusa peruviana</i>	4	158	3	165	0.71
Glyceridae	<i>Glycera americana</i>	16	15	8	39	0.17
...						

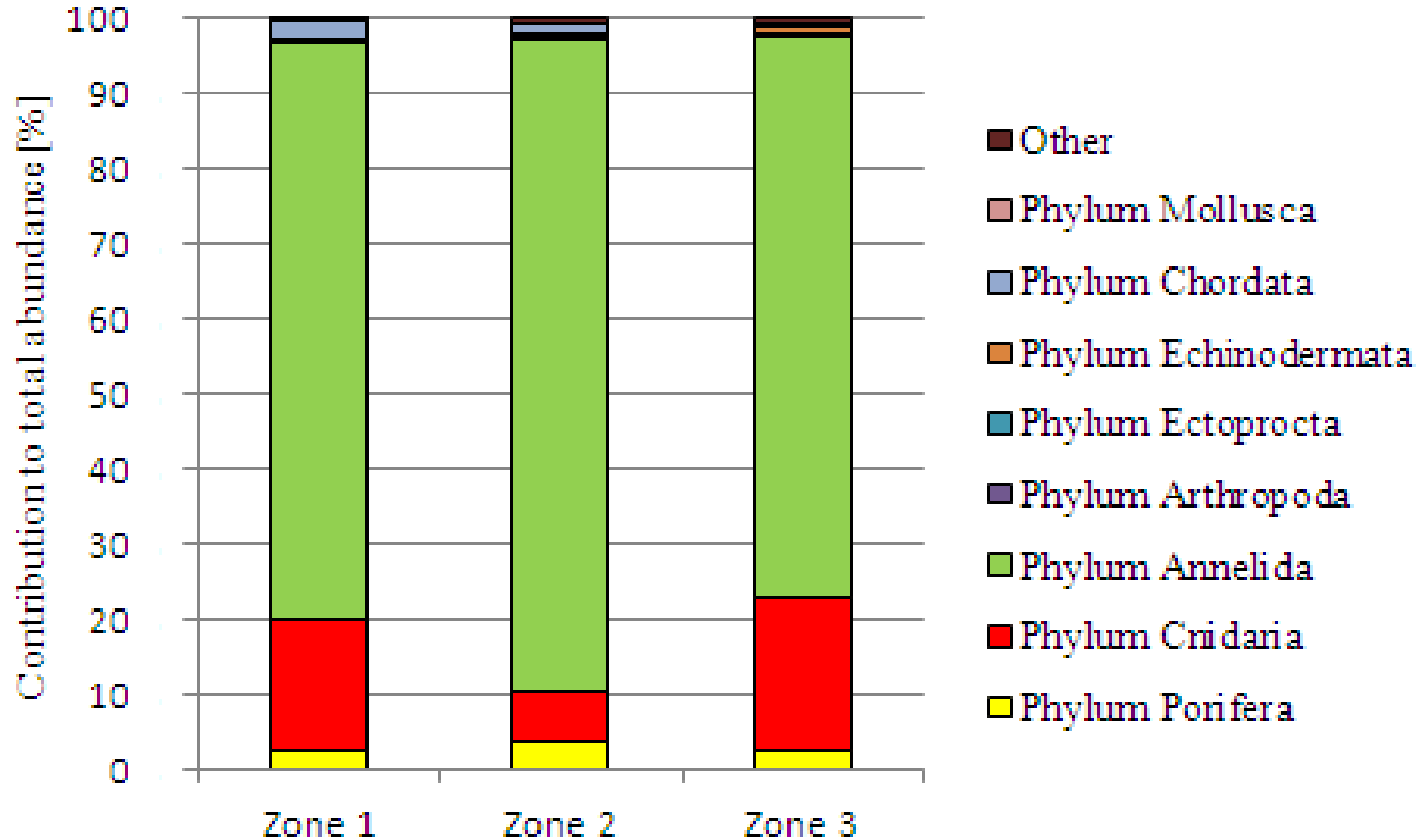
Community Description

Restsediment



- Abra alba*
- Astarte borealis*
- Corbula gibba*
- Modiolarca subpicta*
- Lagis koreni*
- Pygospio elegans*
- Terebellides stroemi*
- Dendrodoa grossularia*
- Arctica islandica*
- Astarte elliptica*
- Kurtiella bidentata*
- Dipolydora quadrilobata*
- Nereimyra punctata*
- Scoloplos armiger*
- Ciona intestinalis*
- Sonstige

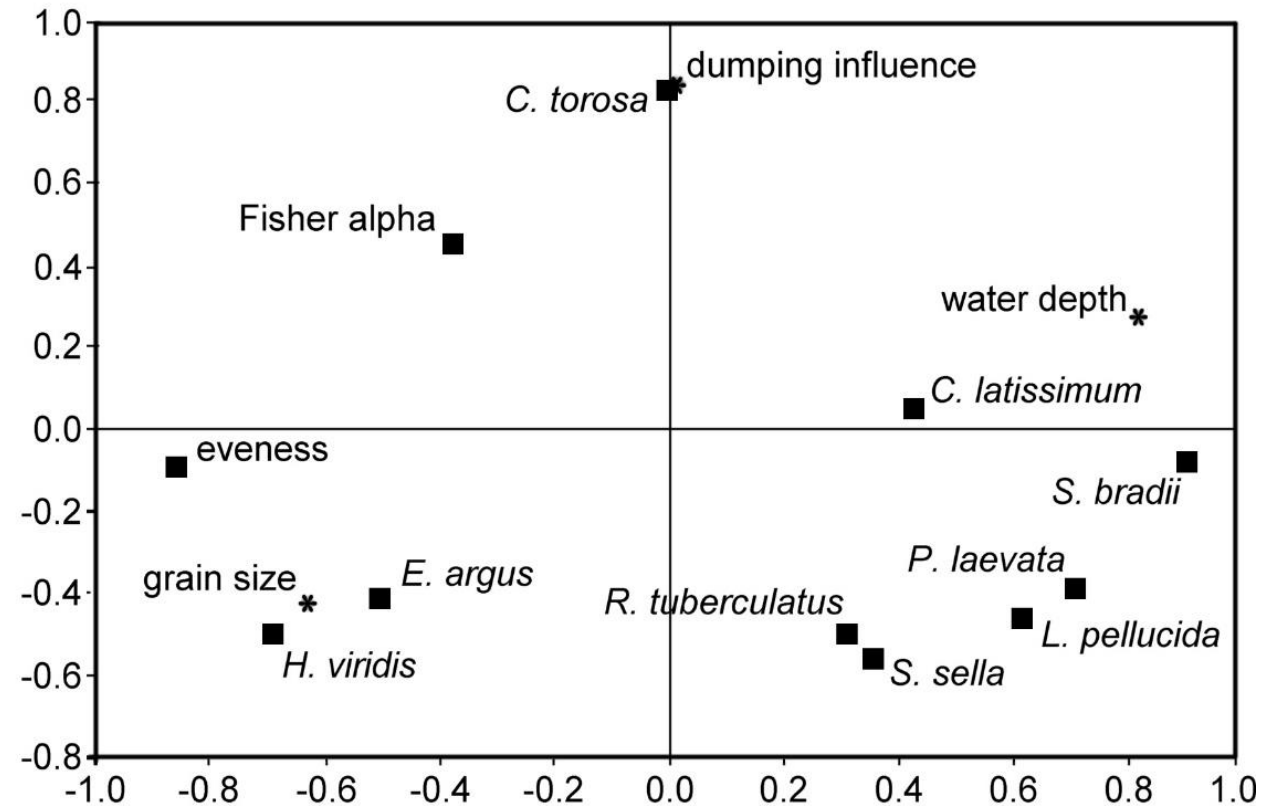
Community Description



Multivariate statistics

- Identification of environmental factors driving distribution of taxa
- Set up of transfer functions

↑ sediment dumping



← turbulence of water

Acknowledgements

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Marine and Fisheries Sciences**



UNIVERSITY OF GHANA
DEPARTMENT OF MARINE AND FISHERIES SCIENCES

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Conclusion

Biological organisms can be use for monitoring water quality due to:

- Sedentary life style
- Species specific sensitivity
- Presence or absence of certain invertebrates is a determinant of the **state of aquatic ecosystems**

